

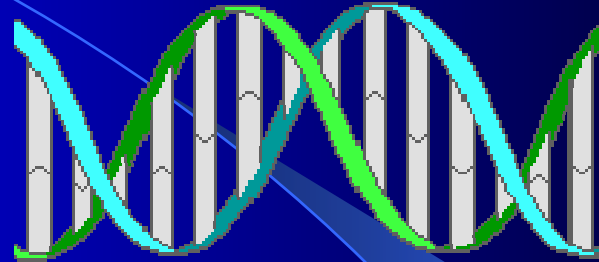
What DNA tells us about Walleye (& other fish) in the Great Lakes



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What do we use DNA data for?



- 1. Delineate stocks
- 2. Determine which spawning groups/locations are the most critical
- 3. Measure changes in genetic diversity over time, in the face of exploitation and irregular year classes and recruitment
- 4. Predict where individuals originated (spawning group)
- 5. Match up eggs, larvae with adult life history stages
- 6. Identify unknowns (i.e., fish fillets, parents of fry, etc.)

Objectives of Our Study



To develop, test, analyze, and implement a high-resolution, low cost, and widely applicable DNA data base for analyzing fish stock structure in the Great Lakes for walleye and yellow perch.

Types of DNA data we are collecting: A Dual approach

Mitochondrial DNA Sequences:

Maternally-inherited
Can see clear geographic and historical patterns, and relation to other species
More expensive
Single locus

Nuclear DNA Microsatellite variation:

Biparentally-inherited
Less expensive, high through-put
Increased resolution power due to multiple loci
Couples well with mtDNA to address a variety of questions

Microsatellite DNA

Microsatellites (or VNTRs = variable number of tandem repeats) are short segments of DNA that have a repeated sequence such as CACACACA, which occur in non-coding DNA.

Microsatellites mutate rapidly and have no known function = “junk DNA”.

These mutations are in the form of losses or gains of repeats.

Individuals in a population typically possess microsatellite alleles of different numbers of repeat copies, having variable lengths.

CACACACACACACACACACA 10
CACACACACACACACACACA 11
CACACACACACACACACACACA 12
CACACACACACACACACACACACA 13
CACACACACACACACACACACACA 14

Inheritance of Microsatellites

CACACACACACACACA 9
CACACACACACACACACACACACA 14

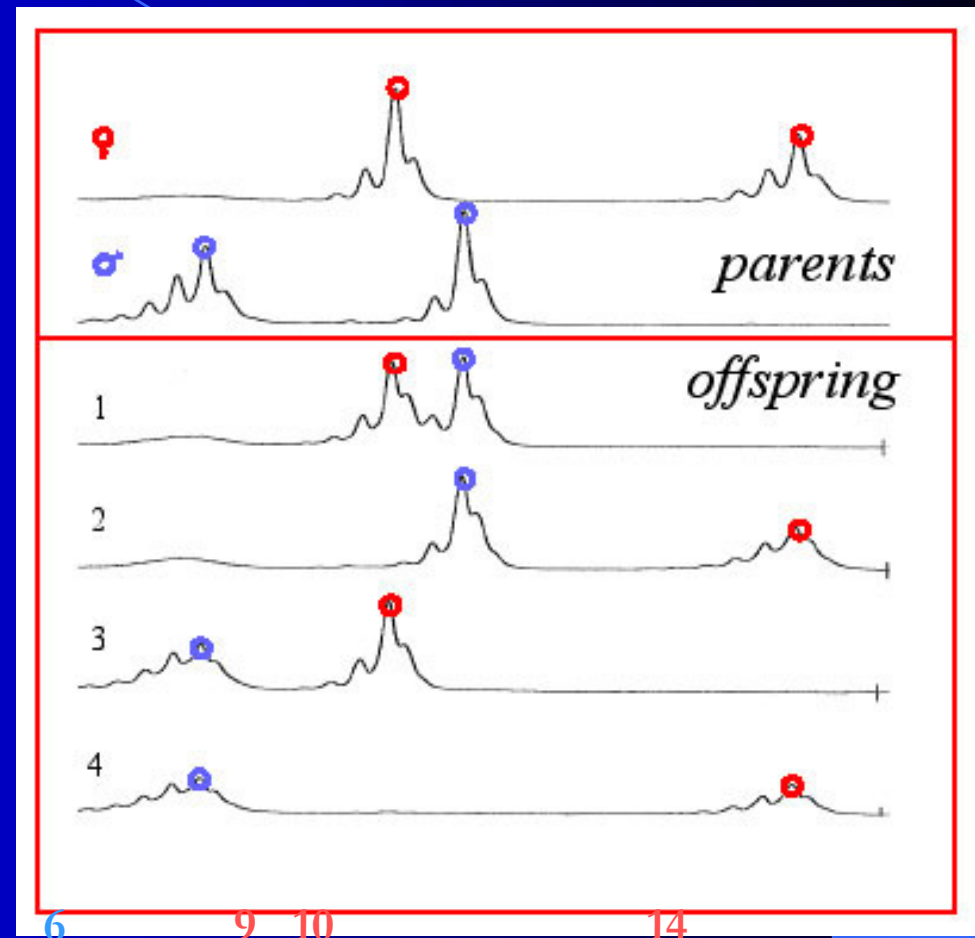
CACACACACACA 6
CACACACACACACACACA 10

CACACACACACACACA 9
CACACACACACACACACA 10

CACACACACACACACACA 10
CACACACACACACACACACACACA 14

CACACACACACA 6
CACACACACACACACA 9

CACACACACACA 6
CACACACACACACACACACACACA 14



Diploid organisms (such as walleye and humans) each have 2 copies.

Populations (stocks) that are isolated diverge in microsatellite frequency lengths over time

	Pop A:	Pop B:
9	10%	1%
10	80%	35%
11	8%	47%
12	2%	17%

We assay several different microsatellite loci to test this hypothesis independently and statistically.

Application of the Study



Our studies build upon the past studies to better understand fine-scale stock structure, allow unknowns to be genetically typed, and to produce a large interactive data base at low cost for use by fishery scientists and managers.

Genetics of the Walleye *Sander vitreus*

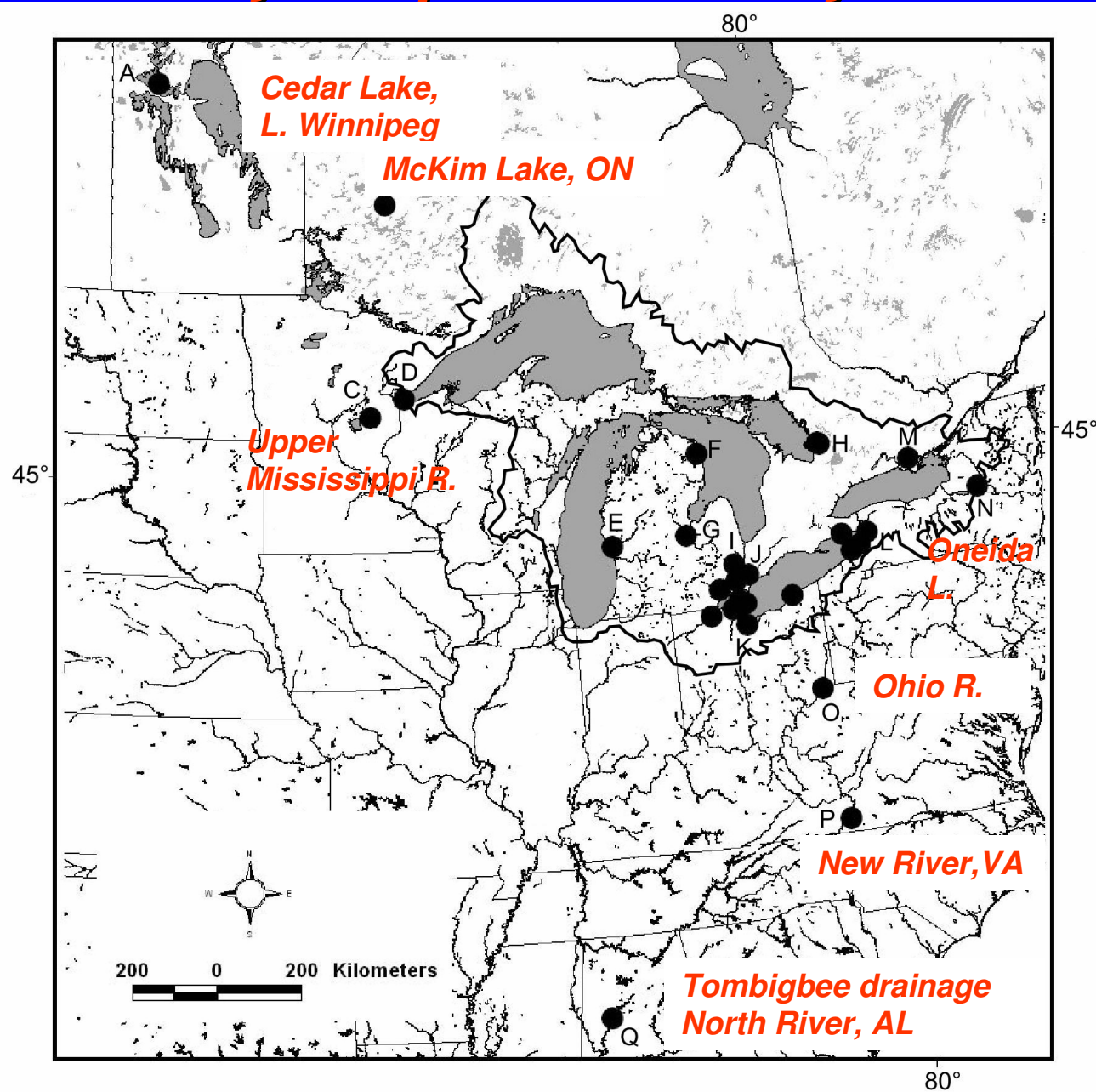


- Our work to date, as well as tagging data & ecological data, indicates that there are significant differences in genetic composition among spawning groups
- These data appear to support spawning site philopatry (i.e., natal homing)
- Differences among populations in the Great Lakes have been maintained by this behavior since their founding after the Ice Ages

Methods

- Analyzed 10 (going to 15) nuclear microsatellite loci
- **1000+ Walleye**
- **28 spawning sites**
 - Great Lakes (Lakes Superior, Michigan, Huron, St. Clair, Erie*, Ontario)**
 - Northwest outlying populations:**
 - Lake Winnipeg area – Cedar Lake**
 - Southwest Ontario -McKim Lake/Papaonga R.**
 - upper Mississippi River drainage - Mille Lacs**
 - Southeast outlying populations:**
 - Ohio River drainage**
 - Tennessee/Tombigbee R. drainage to Mobile Bay, Gulf of Mexico (North River)**

Walleye Population Study Sites

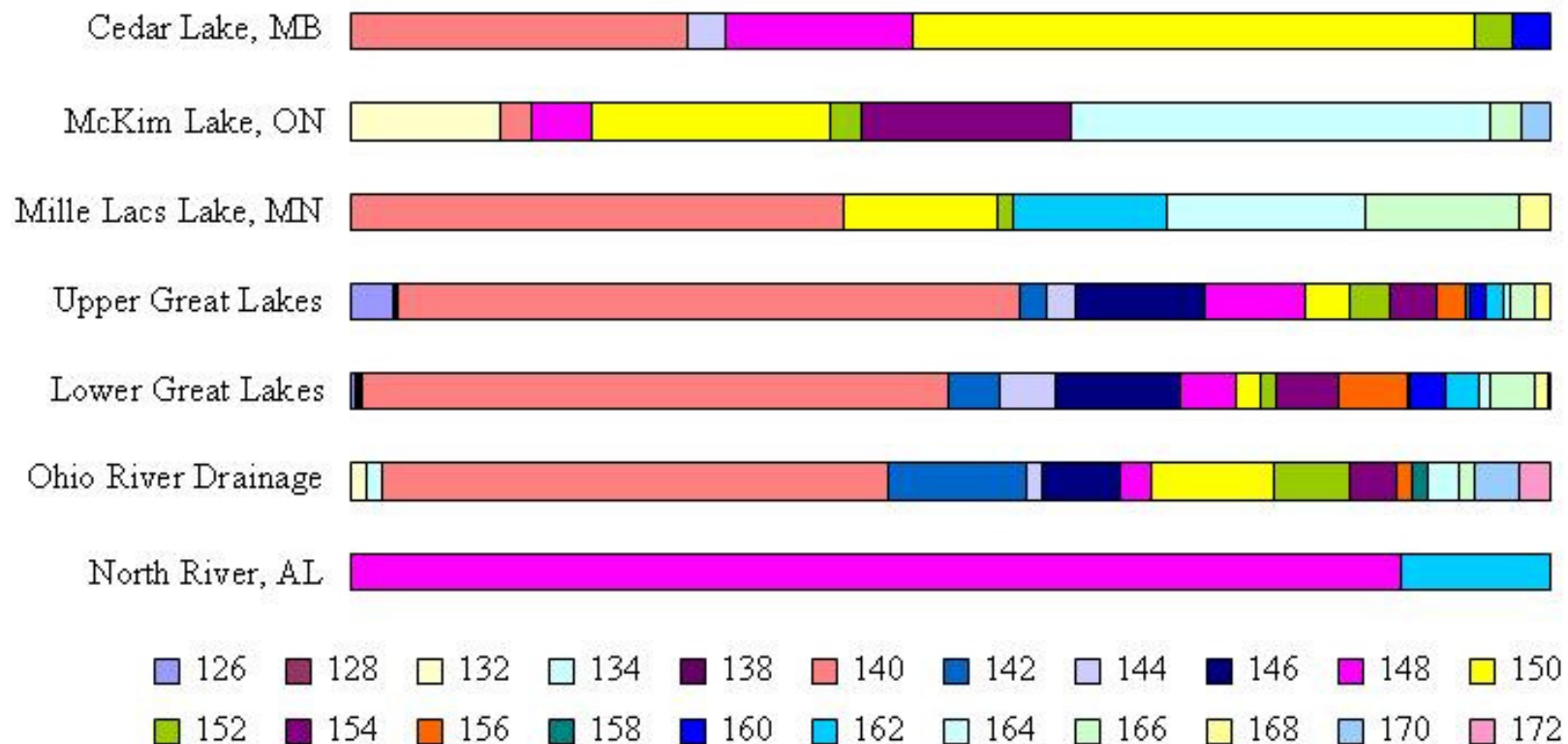


Genetic Diversity Comparisons

Site	<i>N</i>	Heterozygosity	Proportion private alleles
Lake Winnipeg	16	.60	.02
Upper Mississippi River	39	.58	.05
Lake Superior	38	.62	.02
Lake Michigan	50	.69	.00
Lake Huron	125	.70	.03
Lake St. Clair	78	.73	.03
Western Lake Erie	319	.69	.03
Eastern Lake Erie	182	.72	.02
Lake Ontario watershed	70	.69	.00
Ohio River drainage	39	.68	.01
Tennessee River drainage	06	.50	.20
Total/Mean	972	.70	--

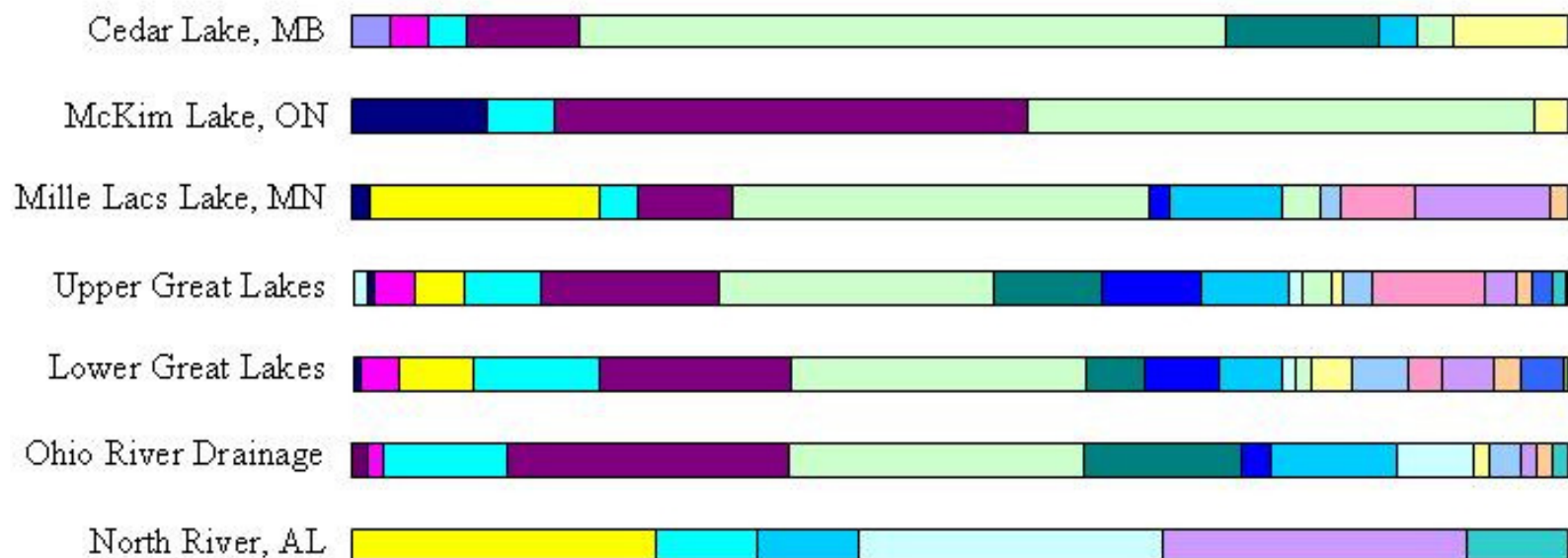
Example: Allelic variation among walleye population sites

Svi 6

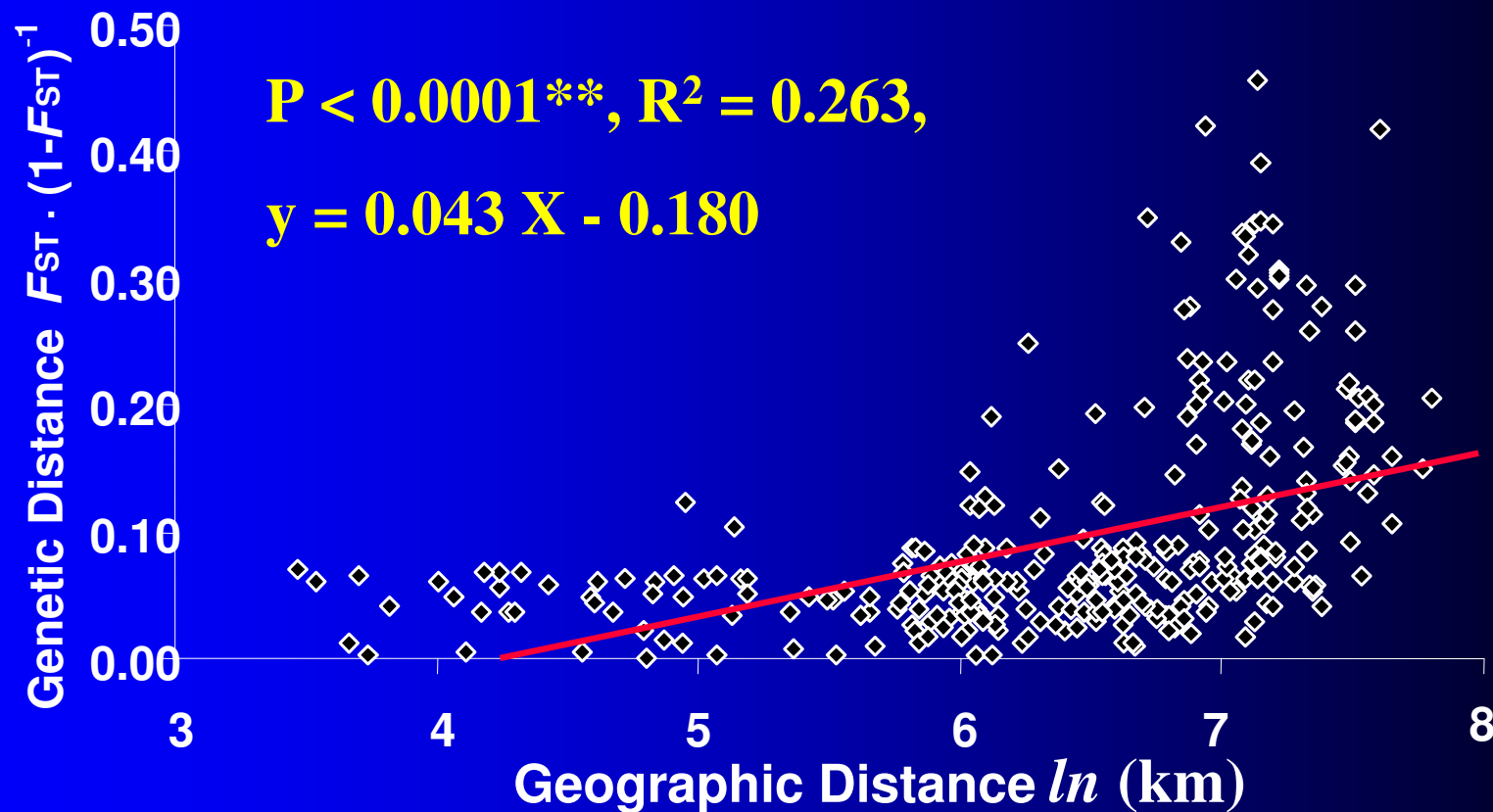


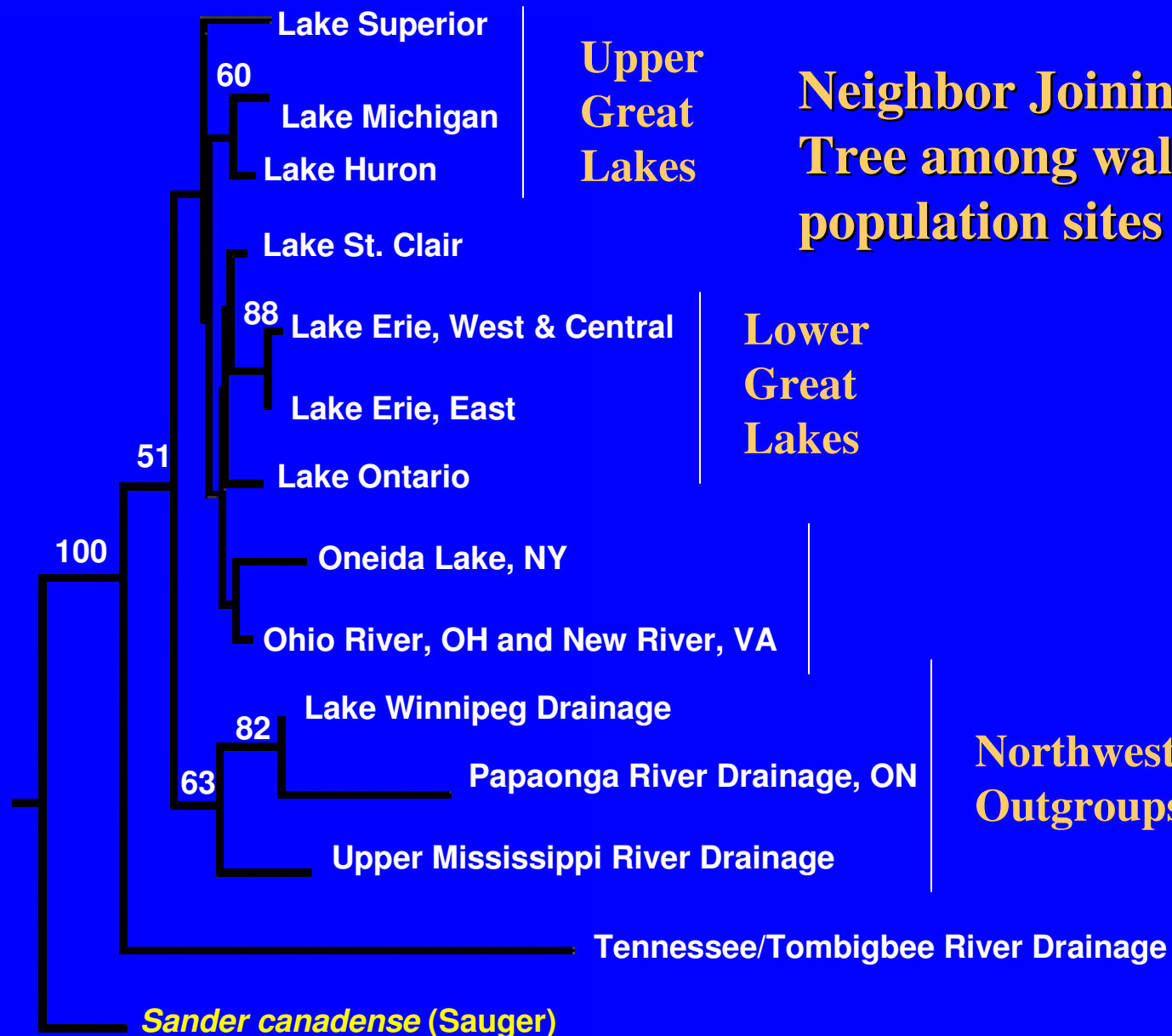
Example: Allelic variation among walleye population sites at *Svi* L7 locus

Svi L7



Mantel Test shows Broadscale Genetic Isolation by Geographic Distance

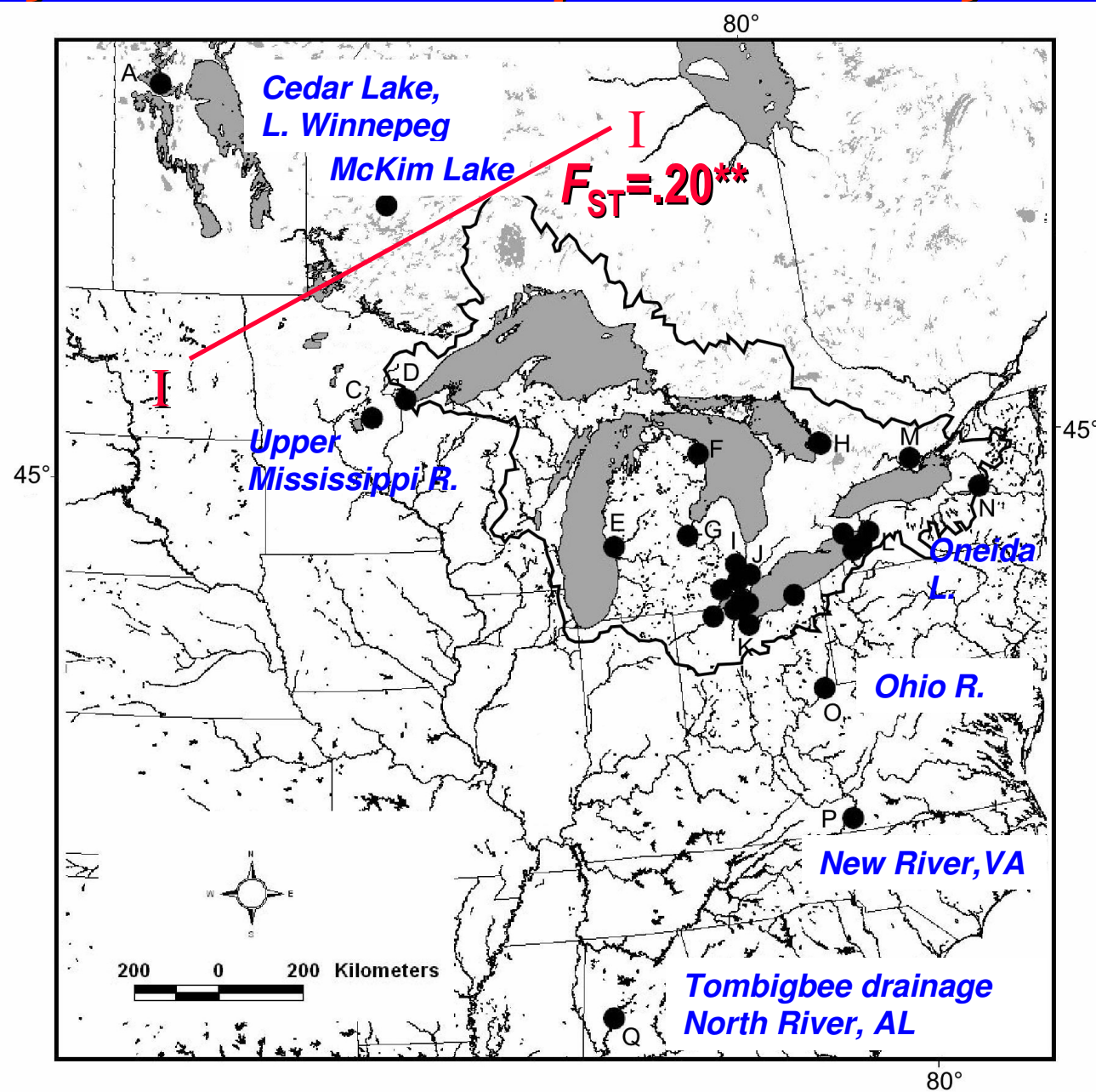




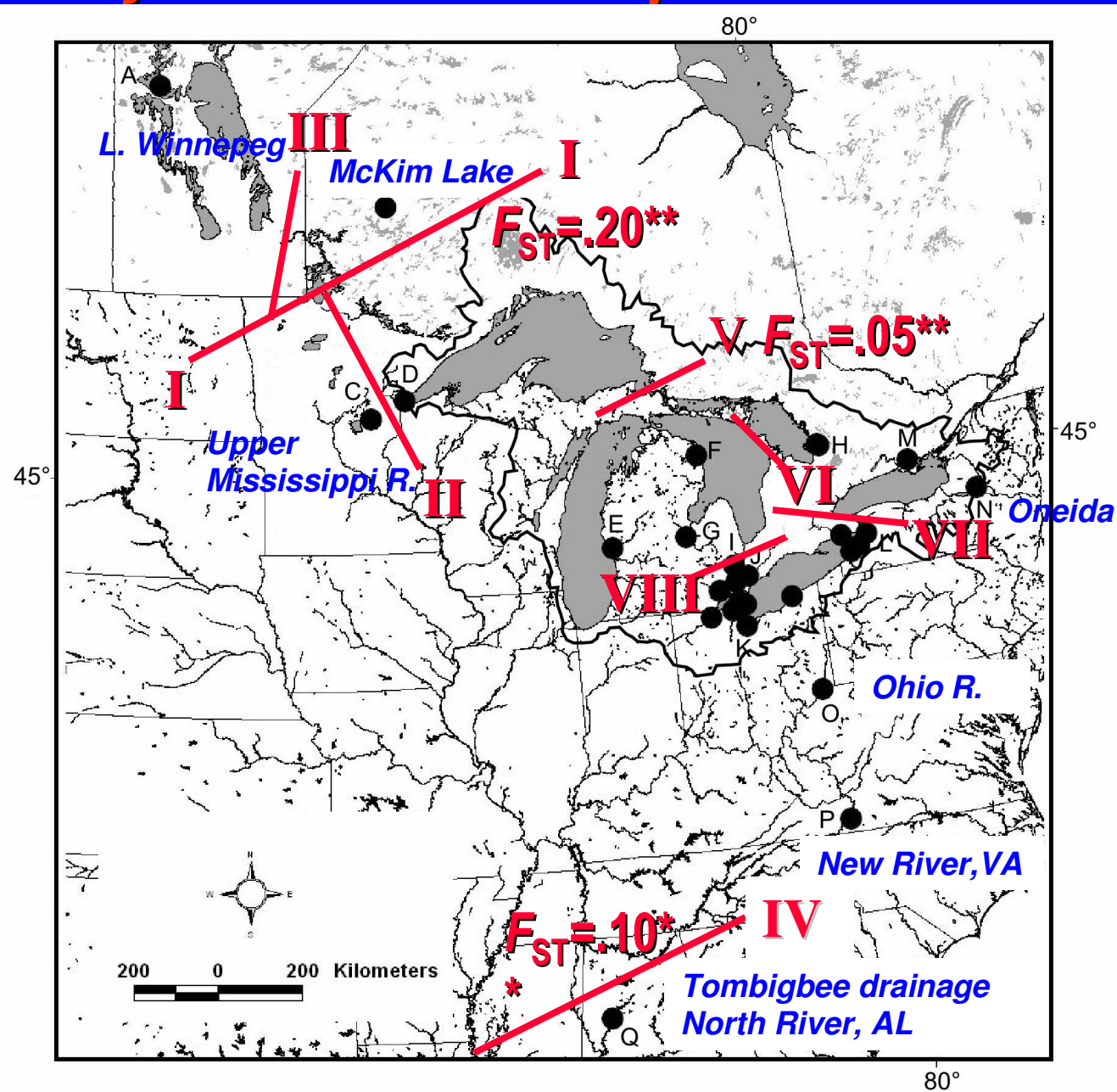
0.01

Nei's (1972) Genetic Distance

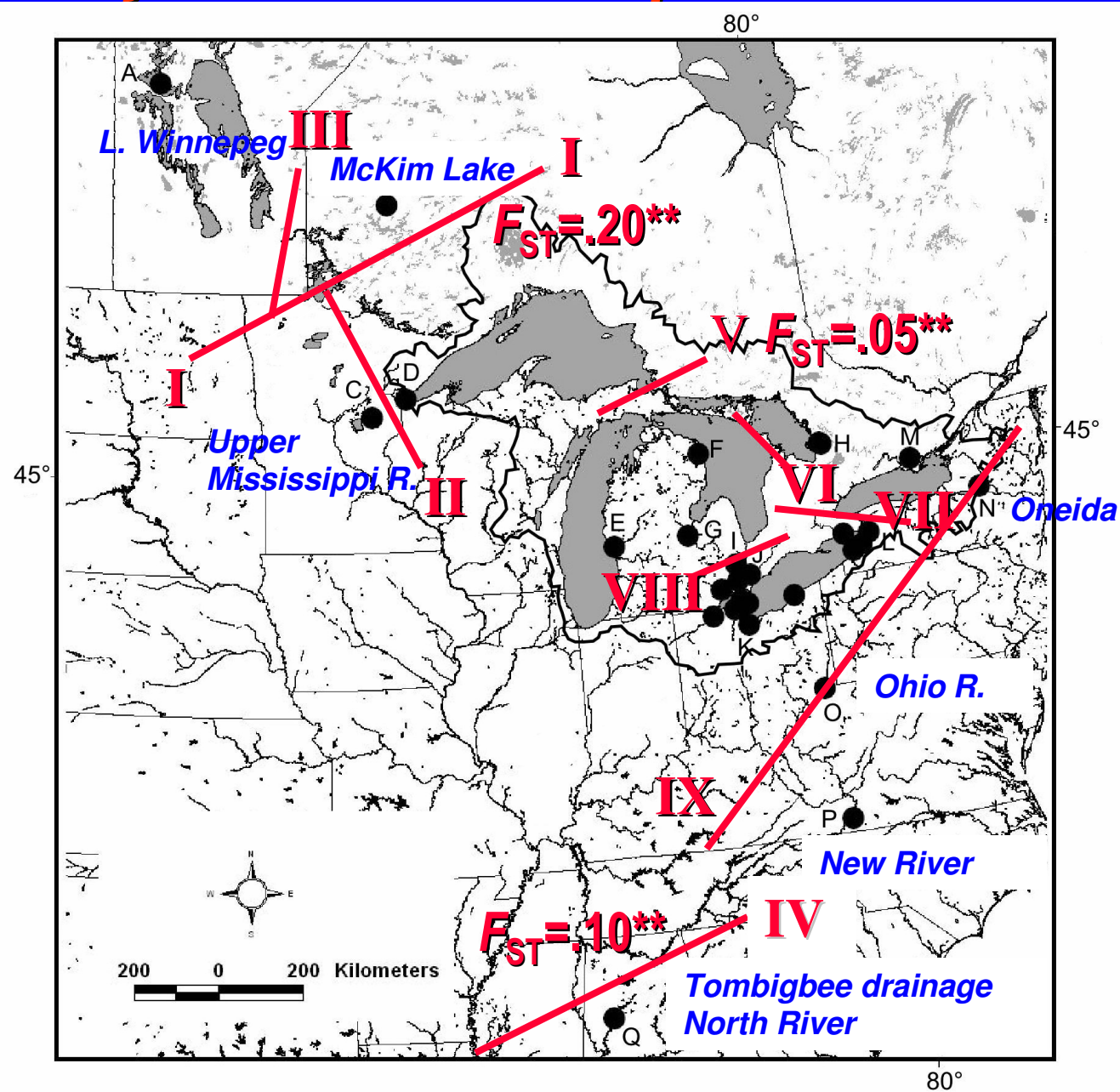
Walleye Broadscale Population Study Barriers



Walleye BROADSCALE Population Barriers

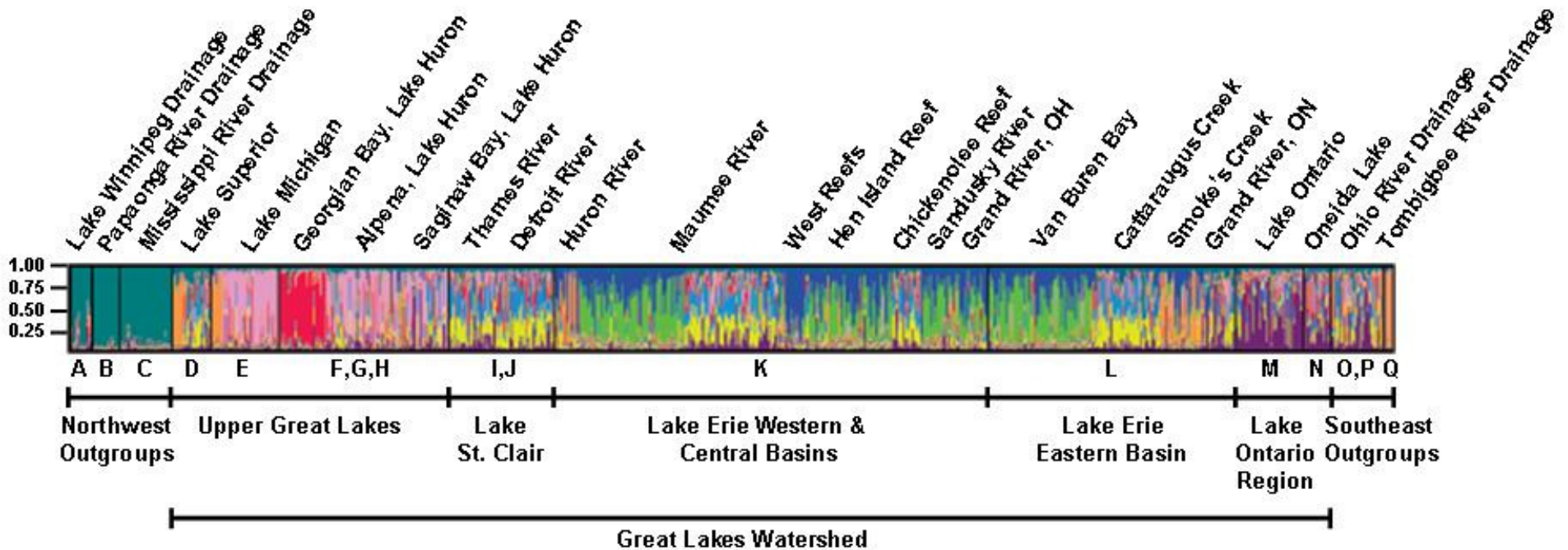


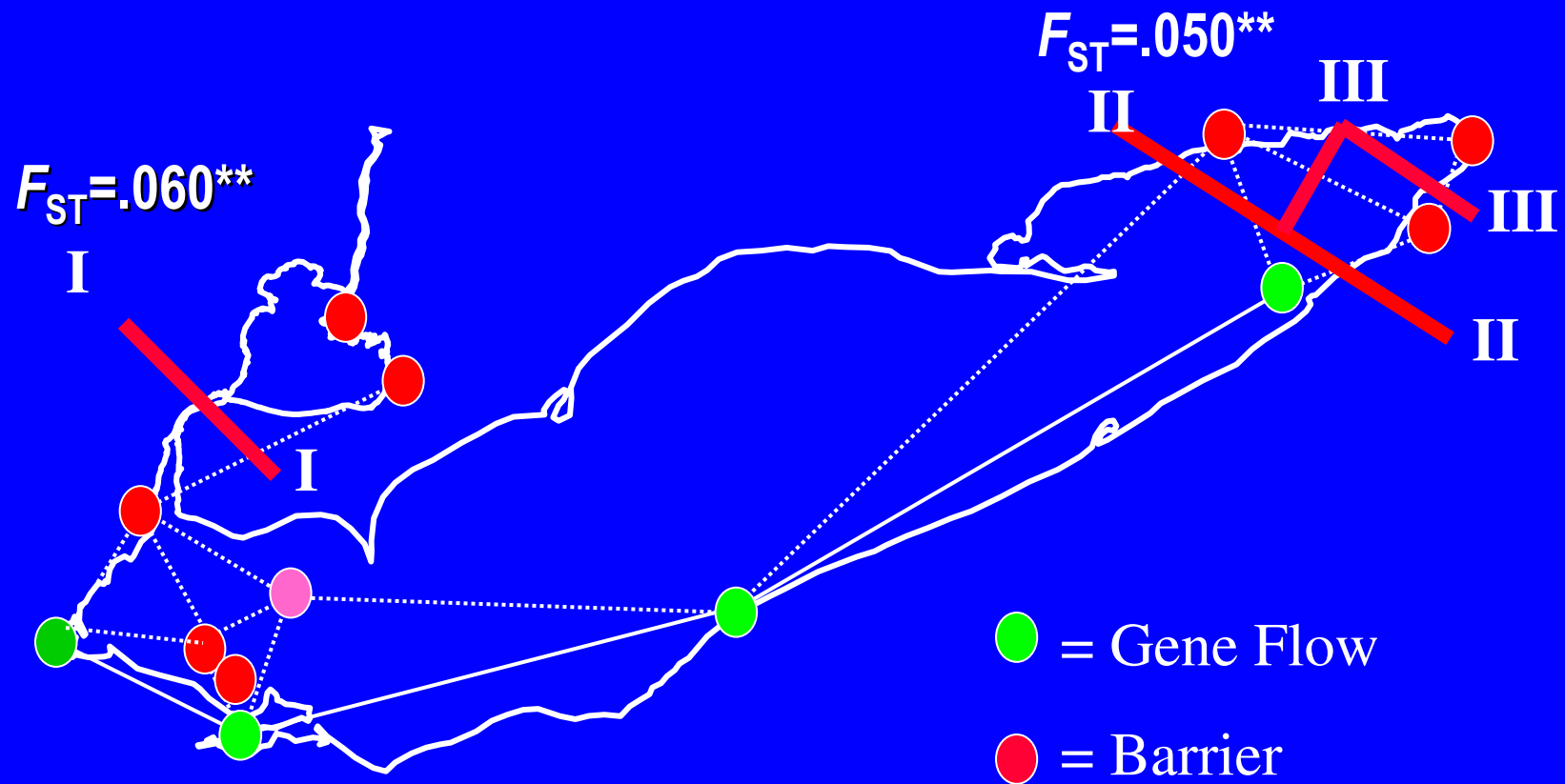
Walleye Broadscale Population Barriers



Bayesian Structure Analysis to Identify Population Groups

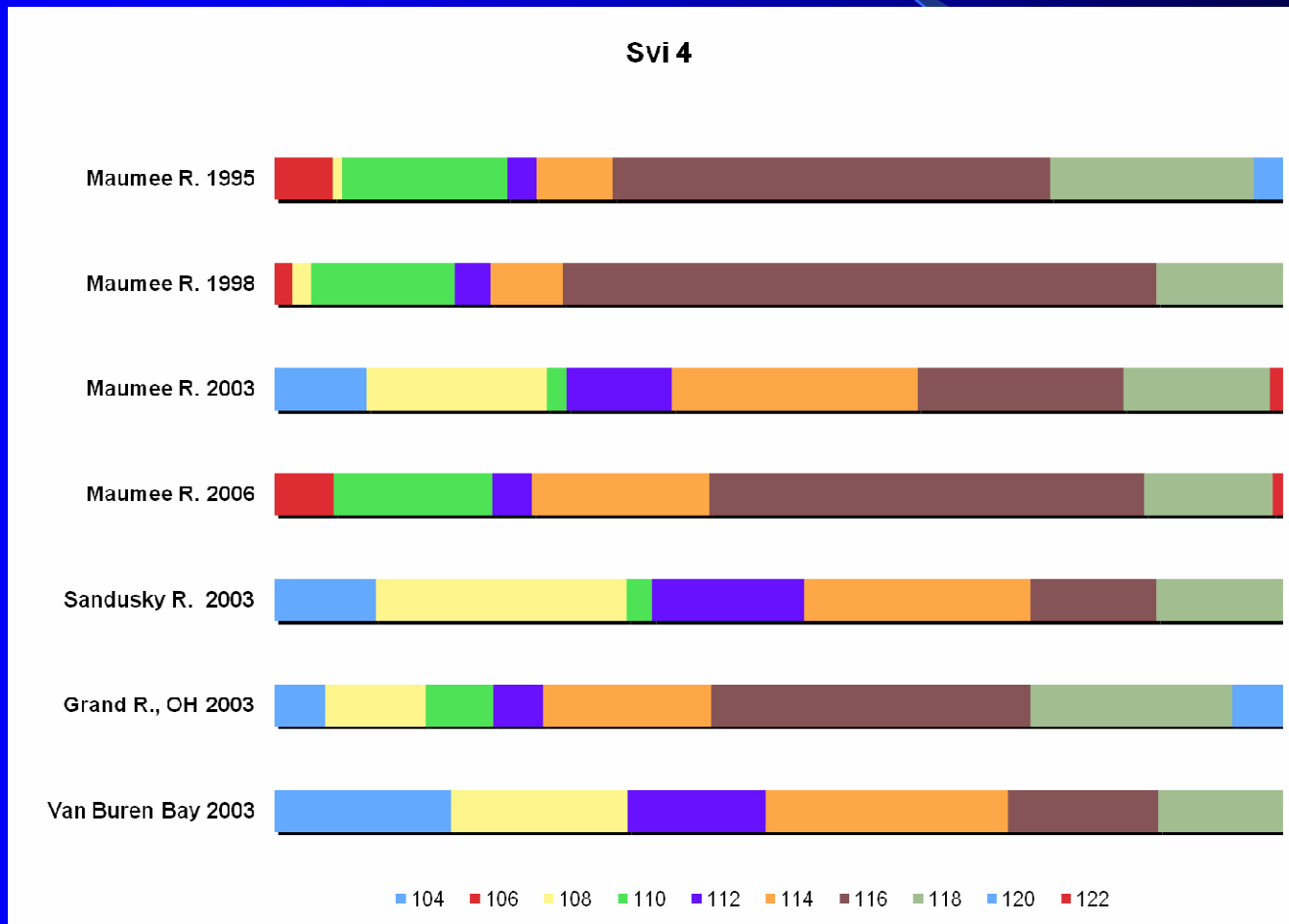
$K=9$, $pp=0.98$





**Walleye Finescale Population Barriers
in Lake Erie $N=501$, 12 Sites**

2003 fine-scale Walleye patterns along Lake Erie southern shore



Differences between Maumee River Walleye run years

	1.	2.	3.
1. Maumee R 1995	~		
2. Maumee R 1998	NS	~	
3. Maumee R 2003	**	**	~
4. Maumee R 2006	NS	NS	**

Conclusions & Summary

1) *Does genetic diversity change across the range of walleye?*

Somewhat, .50-.74; highest in Great Lakes

Is it higher in nonglaciated areas?

No, highest in areas where glacial refugia meet in G. Lakes

2) *How many primary walleye population groups occur across the native range?* 9

3) *What/where are the primary genetic barriers?*

- NW region in Canada (Missouri refugium)
- Mobile Bay drainage, L. Superior, Georgian Bay (L. Huron),
- L. Ontario, L. St. Clair, Ohio R. drainage
- In L. Erie: eastern river sites

4) *Do their genetic patterns fit an isolation by geographic distance hypothesis?* Yes broadscale

No finescale (Lake Erie)

5) *What fine scale patterns are discerned?*

- E Lake Erie walleye very different from W
- Spawning groups mostly are temporally stable
- Some high gene flow years may be linked to higher recruitment

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USEPA

Lake Erie Protection Fund

Thank You!

Collections:

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